

# DIFFERENCES IN CHEMICAL COMPOSITION AMONG PROVENANCES OF BROWSE SPECIES IN A SUBHUMID ENVIRONMENT: RELATION TO USE AS SUPPLEMENTS

B.H. Dzowela<sup>1</sup>, L. Hove<sup>2</sup> and P.L. Mafongoya<sup>3</sup>

<sup>1</sup> SADC-ICRAF Agroforestry Project, P.O. Box CY 594, Causeway, Harare, Zimbabwe

<sup>2</sup> Makoholi Research Station, P/Bag 9248, Masvingo, Zimbabwe

<sup>3</sup> Agronomy Institute, Department of Research and Specialist Services, P.O. Box CY550, Causeway, Harare, Zimbabwe

## ABSTRACT

Studies were initiated in a subhumid Southern African environment to assess the chemical composition and nutritive value of *Acacia angustissima* (Mill.) Kuntze, *Calliandra calothyrsus* Meiss. and *Leucaena* species. The objective was to determine variability in crude protein (CP) concentration, acid detergent fibre (ADF), neutral detergent fibre (NDF), and soluble and insoluble proanthocyanidins from fodder samples of species, subspecies and provenances. A wide range was found in these parameters. The implications of these chemical composition factors, especially proanthocyanidins, on the use of these browse fodders in livestock feeding systems are discussed.

## KEYWORDS

Tree fodders, crude protein, fibres, proanthocyanidins, digestibility, supplements, feeding systems.

## INTRODUCTION

Williams *et al.* (1976) suggested five basic requirements for a good forage species: high dry matter yield, persistence, adequate feed quality, compatibility with other species and ease of propagation and establishment. While a majority of species tested over the years, in Africa (Thomas and Sumberg, 1995) have shown high fodder production potential and adaptation to local edaphic and climatic conditions, socio-economic constraints associated with communal land-tenure and poor livestock marketing infrastructures limit their wide-scale adoption. One of the biophysical limitations to the wide-scale adoption of herbaceous legume fodders is a lack of persistence in smallholder-based production systems (Dzowela, 1993). Instead, the planting of leguminous tree fodderbanks has been promoted through agroforestry and silvopastoral systems (Atta-Krah, 1993) in the subhumid environments. Leguminous trees produce high quality fodder. However, one of the major constraints is their chemical composition and nutritive value, and how these characteristics affect their use in feeding systems. Factors contributing to high quality forage, are high concentrations of N and low concentrations of lignin and polyphenolic secondary plant compounds. Reeves (1985) has found substantial amounts of protein in fibre fractions as cell wall proteins which are bound to lignin and are of limited digestibility by rumen microbes. Thus, the chemical composition of fodder from tree fodders does have a strong influence on its use (Ndlovu *et al.*, 1996). The present study was undertaken to assess the levels of chemical composition variability in multipurpose fodder tree species and provenances so as to relate this variability to potential use as feeds.

## MATERIALS AND METHODS

Three evaluation studies were carried out involving species and provenances in (1) *Leucaena* genus, (2) *Calliandra calothyrsus* Meissen. and (3) *Acacia angustissima* (Mill.) Kuntze. grown at Domboshawa Agroforestry Research site in subhumid Zimbabwe (31° 13' E; 17° 30' S; altitude 1530 m; long-term annual rainfall 895 mm; soils an Haplic livisols).

In the first experiment, the different species and provenances were planted out in fodderbanks. Plots consisted of 3-rows spaced 1.5 m

apart, 15 m long and 4.5 m wide. Treatments were arranged in a randomised complete block design with 4 replicates. Within the row, the plants were planted at 0.5 m spacing. The different species of one genus (*Leucaena*) were planted separately from the other trials, involving *Calliandra* and *Acacia*., fodders from the three tree species were sampled (leaf and stem 10 mm diameter) from 5 randomly selected plants from the middle row of the plot. The fodders were separated into leaf (leaflets and petioles) and stem components, sun-dried for 2-days and ground to pass through 1 mm screen. The forage components were analyzed for the concentration of dry matter (DM) by drying at 105°C for 24 hours; samples for determination of acid detergent fibre content (ADF), neutral detergent fibre (NDF) (Robertson and van Soest, 1981) and crude protein (AOAC, 1990) were oven-dried at 55°C for 48 hours. The content of proanthocyanidins (condensed tannins) was assayed by the butanol-HCl method of Porter *et al.* (1986) while the soluble tannins were determined by the method of Reed *et al.* (1985). In vitro dry matter digestibility (IVDMD) was determined by the Tilley and Terry method (1963).

In the second experiment, DM and nitrogen rumen degradation after 24 and 48 hours of incubation in the rumen, were measured using the nylon bag technique (Orskov *et al.*, 1980). In addition, the water soluble DM and N-fractions of the fodders were determined. Nutrient intake, digestion and N-balance were measured in the 3rd experiment using sixteen 28-month old goats (average weight 26 ± 4.2 kg). The digestion trial involving 4 goats per treatment, was conducted over a 21-day period which consisted of 14-day adaptation and 7-day data collection periods. The goats were fed natural grass hay *ad libitum* and all three *Leucaena* fodders were offered at the rate of 120 g/hd/day.

All data were subjected to an analysis of variance using the procedures described by Steel and Torrie (1980).

## RESULTS

**a) *Leucaena* species and provenances evaluation:** The comparison of nitrogen contents between *L. leucocephala* cv. Cunningham (LL), *L. diversifolia* subspecies *stenocarpa* cv. OFI 53/88 (LD) and *L. pallida* (LP) cv. of CPI 58980 (syn. *L. esculenta* subspecies *paniculata*), showed the contents to be high (Table 1). However, 50% or more of this N was bound to NDF. All three species contained substantial amounts of insoluble proanthocyanidins (PAS) which were much higher than that of veld hay.

The chemical composition of other *Leucaena* genus are summarized in Table 2. There was variability in crude protein level; range of low values was around 17.0% in *L. diversifolia* subsp. *diversifolia* and *stenocarpa* to the high in *L. leucocephala* (27.6%), *L. salvadorensis* (25.6%) and *L. shannonii* subsp. *magnifica* (24.5%). Among the species with low levels of NDF were *L. leucocephala* hybrids with *L. diversifolia*, *L. diversifolia* subsp. *stenocarpa* and *diversifolia* with NDF values below 40%. Among the highest were *L. shannonii* subsp. *magnifica*, *L. pallida* and one *L. diversifolia* provenance subsp. *diversifolia* (OFI 45/87). Similarly, a great deal of variability was observed in PAS concentration between species and subspecies and

provenances. The highest values were in *L. pallida* and the two *L. diversifolia* subsp. *stenocarpa* provenances (OFI 35/88 and OFI 53/88). When three contrasting *Leucaena* spp. (*L. diversifolia*, *L. pallida* and *L. leucocephala*) were fed as supplements to low quality native grass hay, there was an increase in DM intake while intake of digestible DM was only increased by supplementing with *L. pallida* and *L. leucocephala*. However, there were no differences ( $P > 0.05$ ) between treatments in terms of digestible NDF intake. Excretion of N was mainly through faeces (over 90 percent) for all diets. Feeding native grass hay alone or with *L. pallida* or *L. diversifolia* resulted in a negative N-balance. Animals fed *L. diversifolia* had lower ( $P < 0.05$ ) urine N relative to the other animals. This is indicative of the protection from rumen microbes of N by proanthocyanidins in *L. diversifolia*. This result is consistent with rumen degradation results (Table 1).

**b. *Acacia angustissima* provenances evaluation:** As shown in Table 3, a wide variability was observed in the parameters considered (CP, ADF, NDF, soluble and insoluble PAS and IVDMD). One of the species *Mimosa scabrella*, introduced from Rwanda's ICRAF Project had the least content of CP., in both the leaf and stem fodder components. Generally, all provenances had higher CP contents in the leaf than in the stem because the leaf does not contain as much structural material as the stem. The range in the leaf IVDMD was from 42% in OFI 65/92 to only 50.9% in OFI 34/88 which was similar to the digestibility coefficients recorded in OFI 66/92 (50.4%) and OFI 68/92 (50.7%). The provenances with higher digestibility in *A. angustissima* were superior to *C. calothyrsus* (41% IVDMD) and not very much lower than *L. pallida* with an IVDMD values of 53.1%.

**c) *Calliandra calothyrsus* provenances evaluated:** From Table 4 it is evident that a wide variability was found in the parameters measured (ADF, NDF, insoluble and soluble PAS and IVDMD). Some provenances, notably OFI 12/91, 61/92 and to a lesser degree 62/92 appear to have digestibility values that are at least 15% higher than the control provenance, OFI 9/89.

## DISCUSSION AND CONCLUSION

From the data on *Leucaena* spp. it is apparent that 3 groups of *Leucaena* are available. The first group is one of low NDF, low to medium PAS and high crude protein concentrations as represented by *L. leucocephala*, its two hybrids with *L. diversifolia*, and *L. shannonii* and *L. salvadorensis*. They represent materials of high nutritive value. Then there is the group which is represented by *L. pallida* and *L. diversifolia* subspecies *stenocarpa* and *diversifolia* to some extent. This group is characterized by high concentrations of PAS with an ABU. value in excess of 45, CP concentration below 20% and NDF values above 35%. These are materials of low nutritive value. The third group is an intermediate group to which the rest of the species, subspecies and provenances belong (including most of the *L. esculenta* species and subspecies). The differences in the world-acclaimed resistance in *L. esculenta* (syn. *L. pallida*) and *L. diversifolia* to the psyllid pest could be related to their condensed tannins content. The efficiency with which these diverse *Leucaena* species and provenances will fit into feeding systems will vary also. Our work has shown lower IVDMD values for *L. diversifolia* and *L. pallida* in addition to higher concentrations of condensed tannin than in *L. leucocephala*. These differences resulted in negative N-balance when the *L. diversifolia* and *L. pallida* were fed as supplements to poor quality native grass hay. As feed intake is positively related to herbage digestibility (Minson, 1990) the data presented here suggest that consumption of the two *Leucaena* forages (*diversifolia* and *pallida*) may be limited by the low digestibility/degradability especially for *L. diversifolia*. Thus, while gains may be realized in

psyllid resistance of *L. diversifolia* and *L. pallida* relative to *L. leucocephala*, losses in livestock productivity resulting from their use in feeding systems must be anticipated. This is especially important where they are incorporated into concentrate rations as substitutes for expensive commercial protein sources. Other studies have shown that feeding of *A. angustissima* and *C. calothyrsus* as hay has problems of low digestibility and rumen degradability (Ahn *et al.*, 1989; Palmer and Schlick 1992; Palmer *et al.*, 1994; Dzowela *et al.*, 1995). In the present study the variability found in chemical composition parameters and IVDMD is interesting. Some of the provenances appear to have dry matter digestibility values that are lower than the generally published data of below 40% for the leaf giving options for improvement through selection. In both species, digestibility of the fodder appears to be strongly influenced by the fibre constituents (ADF and NDF) and the concentration of insoluble proanthocyanidins. Regression analyses in our study have established a high negative relationship, in which the IVDMD (%) relationship with chemical composition parameters in both *Acacia* and *Calliandra* provenances are defined by the regression equations:

$$\begin{aligned} \text{IVDMD \%} &= -0.340 (\text{DM \%}) - 0.941 (\text{NDF \%}) \\ (\text{Acacia}) &= -0.949 (\text{ADF\%}) + 0.623 (\text{ins. PAS}) + 0.160 (\text{sol. PAS.}) \\ R^2 &= 0.94 \text{ ***} \\ \text{and;} & \end{aligned}$$

$$\begin{aligned} \text{IVDMD (\%)} &= -0.141 (\text{DM \%}) - 0.928 (\text{NDF \%}) - 0.935 (\text{ADF \%}) + \\ (\text{calliandra}) &= 0.576 (\text{ins. PAS}) + 0.739 (\text{sol. PAS}). \\ R^2 &= 0.90 \text{ ***} \end{aligned}$$

The effect of tannins in fodder legumes on intake of cereal crop residues is important for feeding ruminants in the subhumid Southern Africa because these fodder legume trees can be used as protein supplements (Reed *et al.*, 1990). Legume trees with high content of PAS are associated with low intakes of crop residue whereas those that contain low to moderate levels of PAS are associated with high intakes (Woodward and Reed, 1989). Low total intake and low true digestibility of protein in turn, affect rate of gain for growing animals. In our studies the use of *L. esculenta*, *L. diversifolia* and *L. leucocephala* as supplements has resulted in different utilization efficiencies with *L. diversifolia* producing lower animal performance than the other two species. From this analysis it is evident that the chemical composition of browse species will affect their potential use in feeding systems. Future research emphasis will be directed to fitting the most promising species and provenances into livestock feed delivery in smallholder production systems.

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**Table 1**

Chemical composition of tree fodders from *Leucaena* species compared to native grass hay and IVDMD, DM (%) and N (%) lost after washing and incubating *Leucaena* forage for 24 or 48 hours in the rumen of sheep.

Constituent	Grass Hay	<i>Leucaena</i> species			SED
		LL	LP	LL	
Dry matter DM, g/kg	950	942	944	943	-
Organic matter (g/kg DM)	950	930	980	940	-
ADF (g/kg DM)	516	272	325	281	-
NDF (g/kg DM)	772	393	459	371	-
N (g/kg DM)	4.3	26.6	18.2	25.1	-
NDI N (%)	74.4	56.4	54.9	47.8	-
Insoluble PAS (ABU at <sub>550nm</sub> /g NDF)	1.2	12.8	10.4	8.0	-
IVDMD %	-	48.0	47.8	52.2	-
Washing loss (readily soluble)	DM N	19.5 3.7 <sup>a</sup>	19.9 10.2 <sup>b</sup>	21.5 8.4 <sup>b</sup>	1.3 1.4
Rumen degradability after 24 hours	DM N	38.1 <sup>a</sup> 13.9 <sup>a</sup>	52.0 <sup>c</sup> 48.4 <sup>c</sup>	41.3 <sup>b</sup> 29.0 <sup>b</sup>	1.5 2.5
Rumen degradability after 48 hours	DM N	39.5 <sup>a</sup> 19.0 <sup>a</sup>	56.0 <sup>c</sup> 57.3 <sup>c</sup>	48.1 <sup>b</sup> 36.9 <sup>b</sup>	0.9 1.1

Figures in the row followed by the same letter are not statistically different at P<0.05.

Where NDI N = Neutral detergent insoluble nitrogen

**Table 2**

Chemical composition differences of fodders from *Leucaena* species and provenances

Species Provenances	ID Nos	DM (%)	NDF (%)	Insoluble PAS <sup>a</sup>	CP (%)
<i>L. esculenta</i> subsp. <i>paniculata</i>	OFI 52/87	89.5	36.5	22.5	22.4
<i>L. esculenta</i> subsp. <i>esculenta</i>	OFI 47/87	90.9	40.0	38.2	23.8
<i>L. diversifolia</i> subsp. <i>diversifolia</i>	OFI 45/87	91.2	41.9	29.5	17.0
<i>L. diversifolia</i> subsp. <i>diversifolia</i>	OFI 46/87	90.2	29.5	26.0	19.4
<i>L. diversifolia</i> subsp. <i>stenocarpa</i>	OFI 53/88	92.0	36.3	48.2	17.6
<i>L. diversifolia</i> subsp. <i>stenocarpa</i>	OFI 35/88	91.2	38.5	51.6	17.4
<i>L. leucocephala</i> cv. Cunningham	-	91.2	34.6	15.9	27.6
<i>L. pallida</i>	CPI 58980 offspring	90.1	40.6	63.6	19.5
<i>L. leucocephala</i> x <i>L. diversifolia</i>	ILCA 15090 (Parent=K743)	90.9	39.3	40.2	22.6
<i>L. leucocephala</i> x <i>L. diversifolia</i>	ILCA 15009	90.9	35.1	25.1	19.6
<i>L. shannonii</i> subsp. <i>magnifica</i>	OFI 58/88	92.2	43.6	3.6	24.5
<i>L. shannonii</i> subsp. <i>magnifica</i>	OFI 19/84	91.4	49.3	5.6	25.1
<i>L. pulverulenta</i>	OFI 83/87	90.9	38.9	27.9	22.6
<i>L. salvadorensis</i>	OFI 34/87	93.0	40.0	18.4	25.6
Maize stover	R215	91.5	87.9	nil	3.4
Means ( <i>Leucaena</i> )		91.1	38.9	21.8	
SD±		±0.89	±4.6	±17.2	±3.4

<sup>a</sup> (ABU at <sub>550nm</sub>/gNDF)

**Table 3**Chemical composition and nutritive value data on *A. angustissima* provenances compared with species of high fodder potential in Zimbabwe

Provenance ID	Fodder Component	Crude Protein (%)	ADF (%)	NDF (%)	PAS insoluble	PAS soluble	IVDMD (%)
65/92	leaf	26.9	31.6	18.1	20.0	2.8	42.0
	stem	4.6	54.7	10.2	12.5	3.4	24.5
66/92	leaf	25.6	37.4	14.2	12.2	1.4	50.4
	stem	3.9	50.0	4.7	13.3	2.8	23.2
68/92	leaf	31.0	28.8	10.3	20.4	3.3	50.7
	stem	9.1	54.6	5.9	11.8	3.0	29.3
70/92	leaf	26.3	30.6	17.6	26.1	2.9	46.4
	stem	16.5	53.7	5.2	13.1	2.5	25.1
34/88 (control)	leaf	23.4	42.2	18.0	26.1	3.1	50.9
	stem	7.1	60.1	2.9	13.0	2.1	30.3
<i>L. pallida</i>	leaf	23.1	40.6	63.6	41.4	4.4	53.1
	stem	9.9	64.9	25.8	12.0	3.0	21.2
<i>Calliandra</i> 9/89	leaf	22.9	30.9	17.2	41.7	11.3	41.0
	stem	13.6	55.0	8.1	13.1	2.5	19.9
<i>L. leucocephala</i>	leaf	27.8	29.5	26.0	NA	NA	NA
	stem	9.3	49.2	17.2	NA	NA	NA
<i>Mimosa scabrella</i>	leaf	8.4	64.7	45.3	NA	NA	NA
	stem	4.0	11.7	68.7	NA	NA	NA
Mean	leaf	23.9	24.6	36.4	26.8	4.2	47.8
	stem	8.7	58.4	70.7	12.8	2.8	24.8
SED±	leaf	6.4	0.6	0.7	0.6	0.04	0.7
	stem	4.3	0.3	0.4	0.3	0.02	0.4
CV(%)		-	3.1	2.7	5.8	2.5	3.9

Insoluble PAS = read at 550nm/gNDF in absorbance units

Soluble PAS = read at 550nm/gDM in absorbance units

NA = not assessed

**Table 4**Chemical composition of *Calliandra* provenances sampled in December 1995, at Domboshawa

Provenance	Component	DM (%)	Crude Protein (%)	NDF (%)	ADF (%)	PAS Insoluble	PAS Soluble	IVDMD (%)
10/91	leaf	94	17	39.7	34.2	44.0	8.9	41.1
	stem	94	11	66.1	54.7	18.2	4.8	34.5
11/91	leaf	92	14	49.1	37.7	17.7	6.3	36.7
	stem	93	9	78.2	68.3	21.2	6.2	24.2
12/91	leaf	93	18	38.2	23.0	54.2	8.3	49.8
	stem	94	10	71.9	64.4	12.5	1.7	31.4
23/91	leaf	93	18	46.5	44.6	49.5	5.1	39.9
	stem	93	9	68.9	57.9	76.0	7.1	29.3
147/91	leaf	93	17	42.6	25.5	42.1	10.2	44.5
	stem	94	8	79.2	69.2	16.8	2.4	20.9
45/92	leaf	93	17	40.4	33.8	53.9	10.0	42.3
	stem	93	10	64.2	52.8	17.1	6.4	31.6
53/92	leaf	94	18	38.2	34.8	56.5	9.9	43.9
	stem	93	11	69.2	62.3	18.8	3.0	30.8
56/93	leaf	94	13	42.0	23.3	33.9	8.3	45.9
	stem	94	10	77.7	63.6	21.0	4.1	26.1
61/92	leaf	93	17	38.6	22.9	41.6	6.9	50.4
	stem	94	10	77.8	66.8	20.4	4.3	22.5
62/92	leaf	94	16	37.2	23.7	47.0	10.2	47.2
	stem	92	10	64.3	53.2	22.3	7.7	38.0
57/93	leaf	94	17	40.8	31.7	54.1	9.2	42.7
	stem	93	10	64.9	53.8	18.2	5.9	35.7
9/89	leaf	95	14	40.9	28.4	41.7	11.3	41.0
	stem	95	7	85.1	72.3	13.1	2.5	19.9
Mean	leaf	94	16	41.2	30.3	44.7	8.7	43.8
	stem	93	10	72.3	61.6	23.0	4.7	28.7
SED	leaf	0.09	2.5	0.65	0.92	1.04	0.04	1.25
	stem	0.04	1.3	0.27	0.38	0.42	0.02	0.51
CV(%)		0.18	-	2.3	4.0	6.14	1.3	6.9

Insoluble PAS = read at 550nm/gNDF in absorbance units

Soluble PAS = read at 550nm/gDM in absorbance units